3D Cell Culture (3DCC) on LifeGel® - characteristics and application in drug efficacy testing, immuno-oncology studies and other biomedical research

Marcin Krzykawski¹, David Earnshaw¹, Joanna Kwiatkowska¹, Renata Krzykawska¹, Krzysztof Klimkiewicz¹, Michal Mikula²,

Malgorzata Statkiewicz², Katarzyna Unrug-Bielawska², Magdalena Cybulska-Lubak², Jaroslaw Skokowski³, Natalia Marek-Trzonkowska⁴, Tomasz Sitar⁵,



Przemyslaw Bielski⁵, Anna O'Leary⁶; Carlos Farkas⁶; Anna Czarna⁷, Justyna Kocik-Krol¹

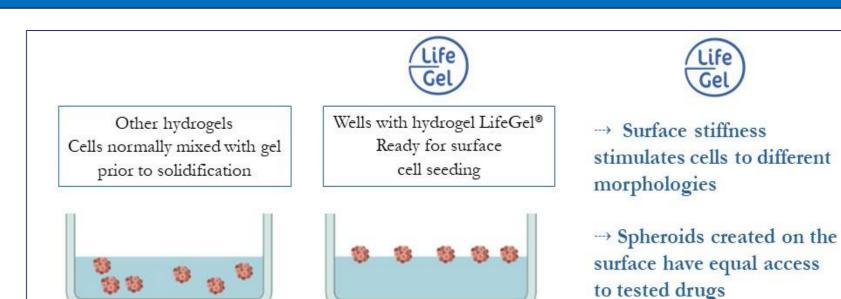
¹ Real Research, Poland, ² Maria Skłodowska-Curie National Research Institute of Oncology, Poland;,

³ St. Adalbert's Hospital, COPERNICUS, Poland, ⁴ International Centre for Cancer Vaccine Science, University of Gdansk, Poland,

⁵ Recepton, Poland, ⁶ Arcella, Netherlands, ⁷ Malopolska Centre of Biotechnology, Jagiellonian University, Poland

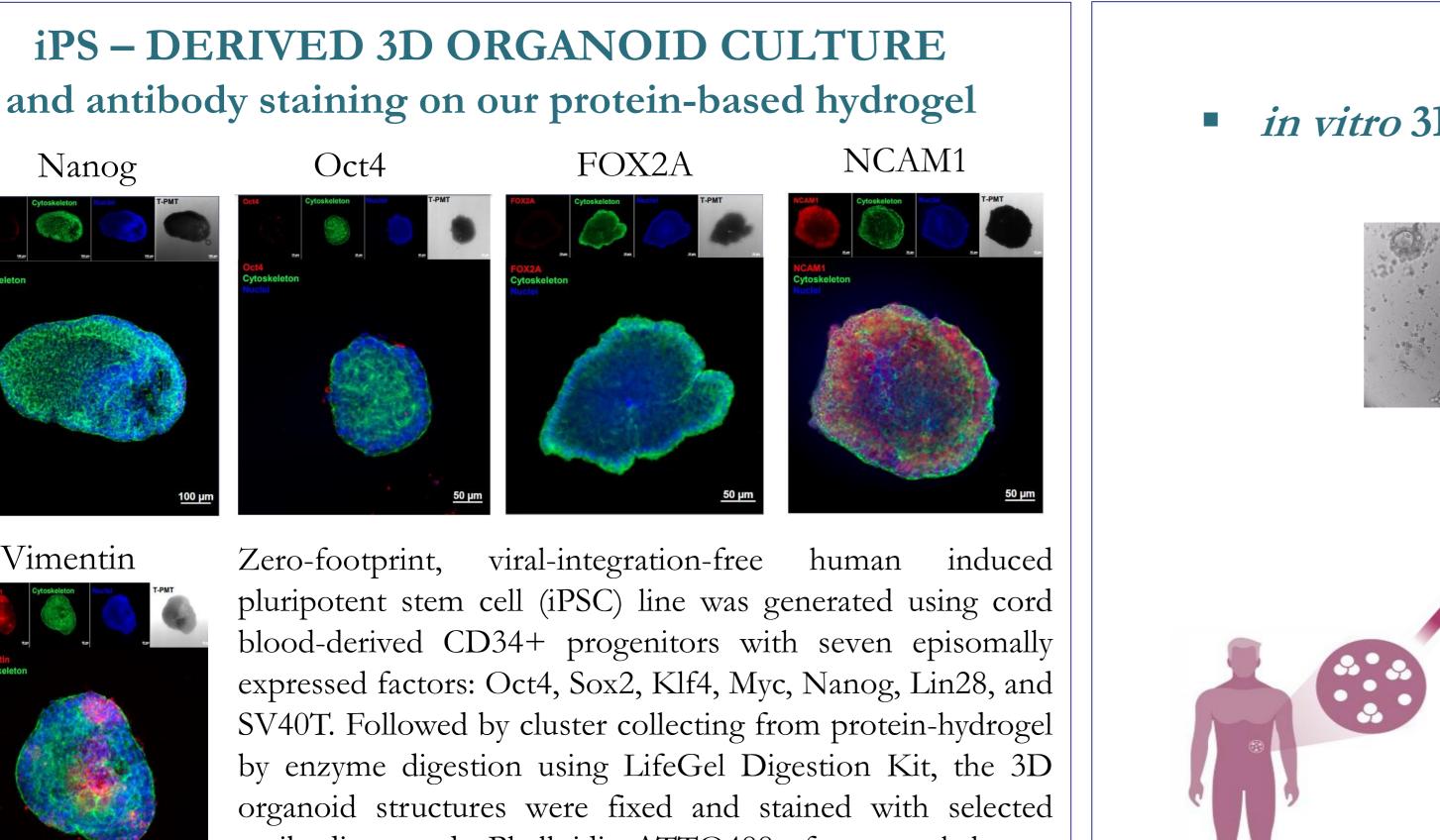
Introduction

In the ever-evolving landscape of biomedical research, 3D Cell Culture (3DCC) systems have emerged as pivotal tools for mimicking in vivo environments and fostering more physiologically relevant cellular responses. It is widely known that 3DCC models may play a crucial role in preclinical studies thus reducing costs of drug testing and avoiding the excessive need for animal models. By providing a biomimetic microenvironment, the hydrogels facilitate the formation of complex cellular structures, enabling reliable representation of tissue architecture and cellular interactions. LifeGels® biophysical parameters can be modified: hardness, density and elasticity, making a solution adapted to every type of organoids and spheroids. The unique characteristic of LifeGel[®] based 3DCC is the phenomena that the 3D structures grow on top of the



hydrogel making them accessible to large molecules like: antibodies, ADC, bi-speciffics, cytokines, CAR-T, oncolytic viruses etc.

3DCC on our protein-based hydrogels – characteristics and application

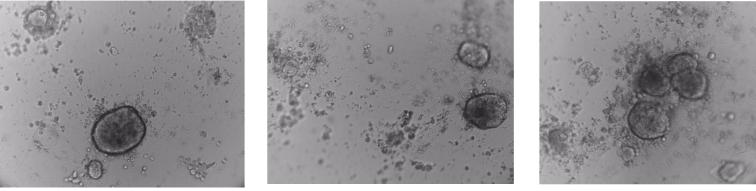


DRUG DEVELOPMENT

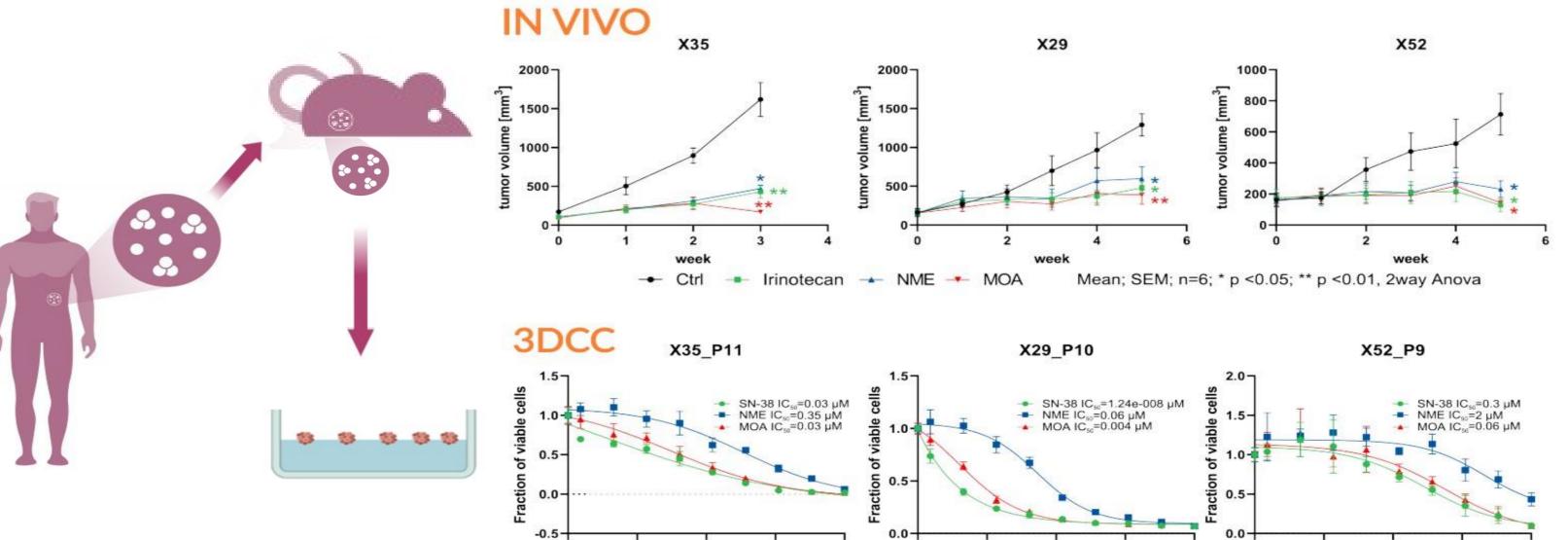
in vitro 3DCC offers reliable predictive models for in vivo outcomes

In collaboration with



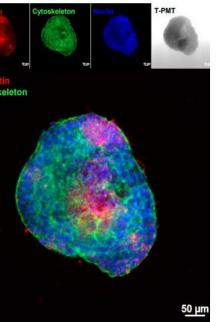


3DCC on of ex vivo culture of PDX-derived cells (colorectal cancer)



Vimentin

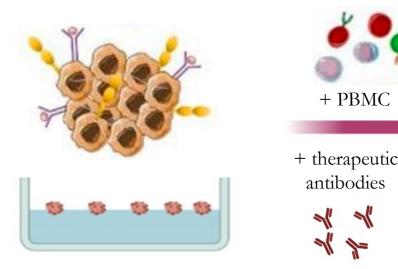
Nanog



antibodies, and Phalloidin-ATTO488 for cytoskeleteon visualisation (of Vimentin) and Hoechst 33342.

3D IMMUNO-ONCOLOGY MODEL

Veryfing of the potency of immune checkpoint inhibitors using 3D co-culture model



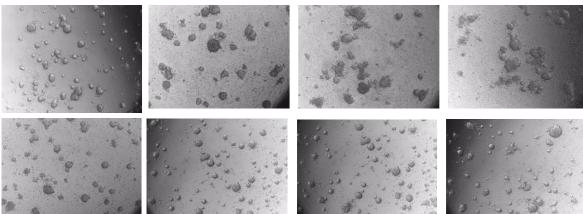
≥ 3DCC surface staining of immune checkpoint proteins on cancer cells.

Mesurement of viability of 3D cancer cells after co-culture with PBMC and antibodies.

3D culture of cancder cells expressing immune checkpoints ligands PD-L1 and VISTA

Cell viability was measured using CellTiter-Blue reagent (Promega) using the protocol optimized for 3D cell culture. The data are presented as a mean value for three PBMC donors. Error bars are represented by SD. * p<0.05, * * p<0.01

culture anti-VISTA anti-CTLA-4 anti-PD-L1 medium



120% iability 40% 20% **3D CULTURE OF CANCER** CELLS + PBMC CELLS ONLY 3D co-culture of cancer cells and PBMC In collaboration with 3D co-culture of • recepton cancer cells only

affinity to science

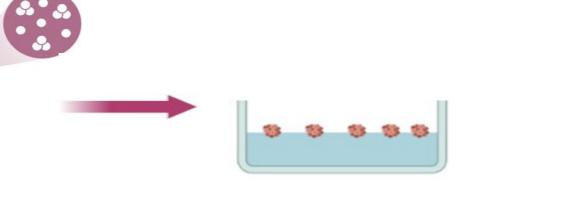
-3

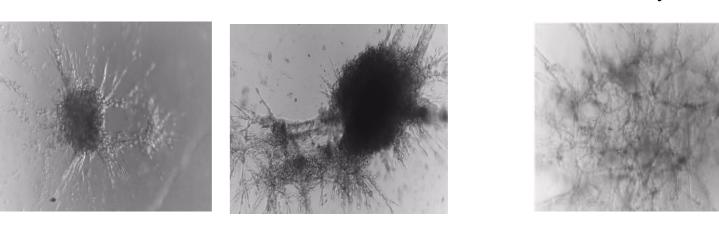
The study demonstrated that the results obtained from 3DCC of PDX-derived colorectal cancer tissues/cells align closely with those observed in *iv vivo* colorecral cancer PDX models, solidifying 3DCC role as a reliable predictive tool of drug response.

in vitro 3DCC of patient-derived cells



3D cell structures of patient-derived non-small cell lung carcinoma cells – different LifeGel® hardness and elasticity



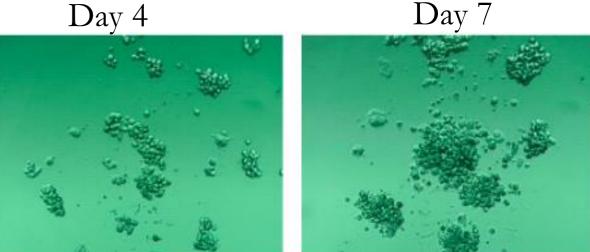


3D Cell Cultures expand drug development

In collaboration with

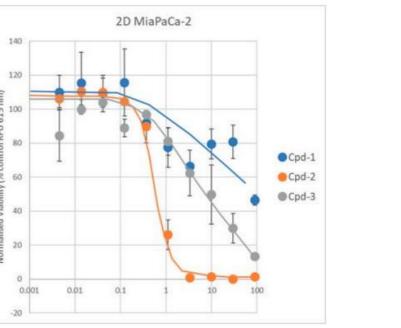


3DCC of MiaPaCa-2 pancreatic tumor cells

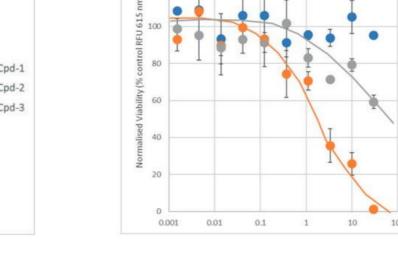








2DCC IC₅₀

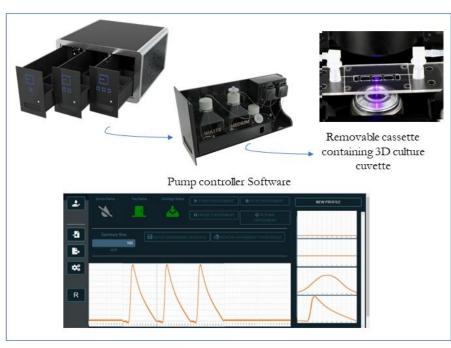


3D MiaPaCa-2





PHARMACOKINETICS STUDIES ON 3DCC

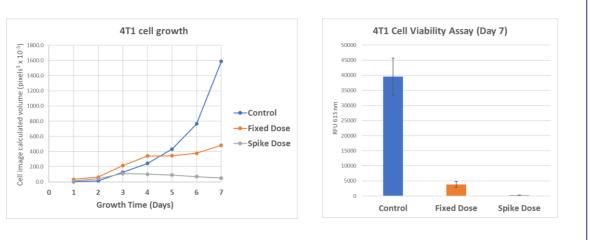


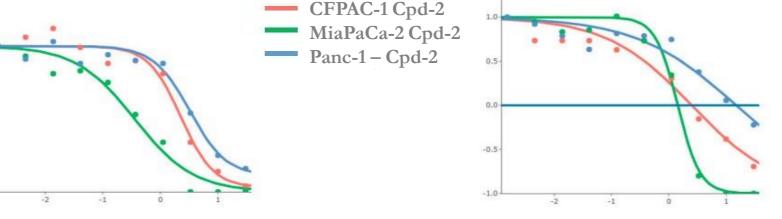
Despite the same total drug quantity being administered, the dose profile may make a significant difference in efficacy, depending on mechanism of action.



Predicting efficacy of different treatment schemes for animals or patients by mimicking PK profiles in the lab.

> Opportunity to measure metabolites and image cell growth in response to different treatment regimes.





	IC 50	GR 50	IC 50	GR 50
CFPAC - 1	2.4	2.6	5.2	0.4
MiaPaCa - 2	0.4	0.2	0.8	0.9
Panc - 1	3.9	3.0	4.9	1.6

Conventional IC50 data for Cpd-2 presents significantly more activity on one of the cell lines in 2DCC, but on 3DCC GR50 parameters for two compounds have similarly high potency, and even the least active is much more active than in 2DCC.

Conclusion

In the realm of cancer research, 3D cell cultures showcase a predictive power in assessing the anticancer effects of drugs that rivals traditional in vivo research models. In our study, we presented a wealth of 3D forms grown on LifeGel® and indicated their use in further stages of 3D organoid culture, drug efficacy testing, immuno-oncology studies and other biomedical research.

Acknowledgement

The research was founded by: Małopolska Centre for Entrepreneurship (MCP) Projects no. RPMP.01.02.01-12-0117/20-00 and RPMP.01.02.01-12.0070/19-00, MCP implements EU funds under the Regional Operational Programme for the Małopolska Region 2014-2020; HORIZON-CSA project CANVAS funded by the European Union, DigiCirc Horizon 2020 project "Right Drug Combination for the Right Colorectal Cancer Patient" funded by the European Union. Polska MAŁOPOLSKA Funded by the European Union

Find us

marcin.krzykawski@real-research.com

